

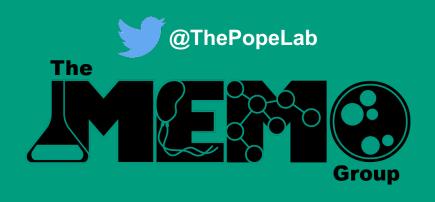
Digital optimization of the feed-microbiome-host nexus

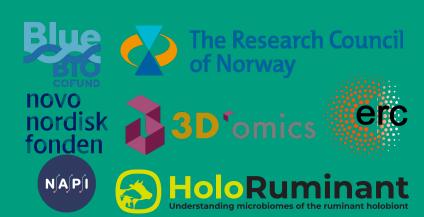
Phil B. Pope + many great MEMO colleagues and collaborators!

Professor

Faculty of Biosciences

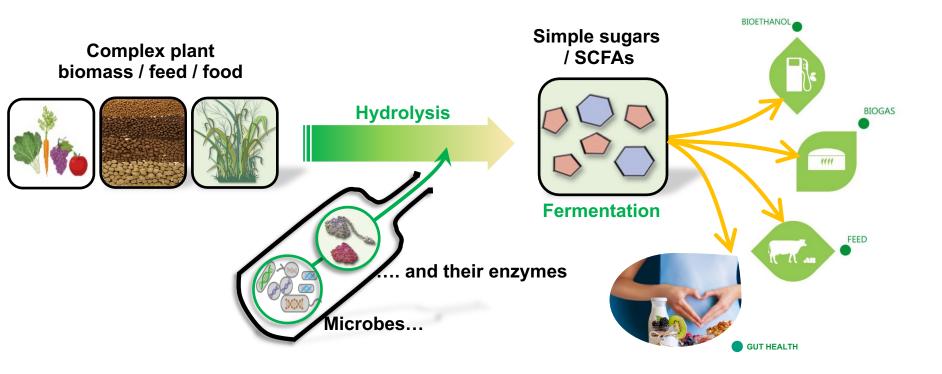
Norwegian University of Life Sciences





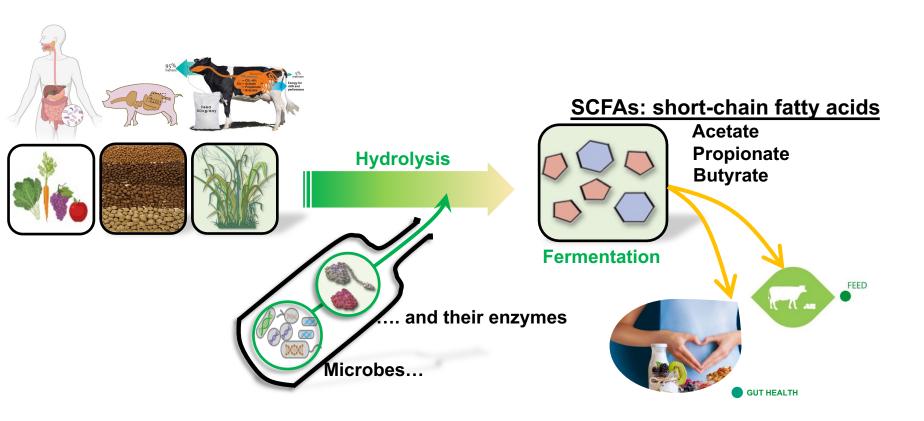


The bottleneck....





The bottleneck....



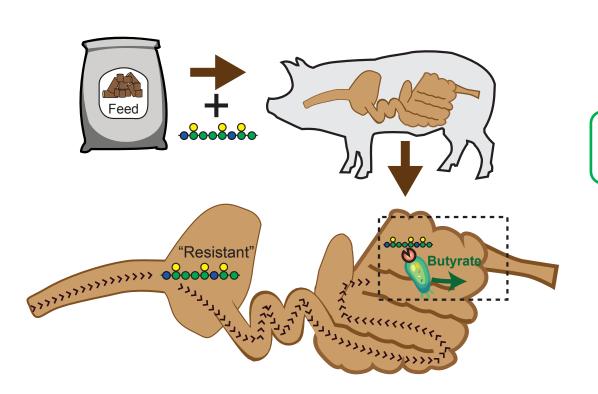
Energy contribution of SCFAs

Cows: ~70%

Pigs: 30–76% Humans: ~10%

The "prebiotic" concept MDF





Common target populations

Lactobacilli Bifidobacteria

Faecalibacterium sp. Roseburia sp.

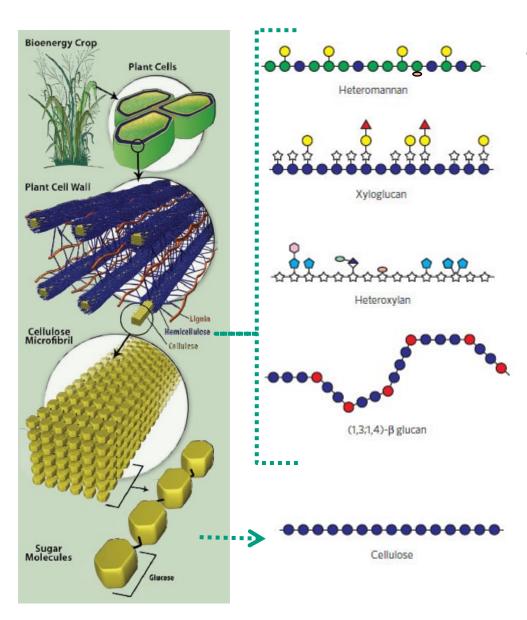




- Healthy gut status
- Low disease state

The plant cell wall





Microbes use: CAZymes

Glycoside hydrolases (GHs)



160+ families

Carbohydrate binding modules (CBMs)



85+ families

Auxilliary Activities (AAs)



15+ families

Carbohydrate esterases (CEs)



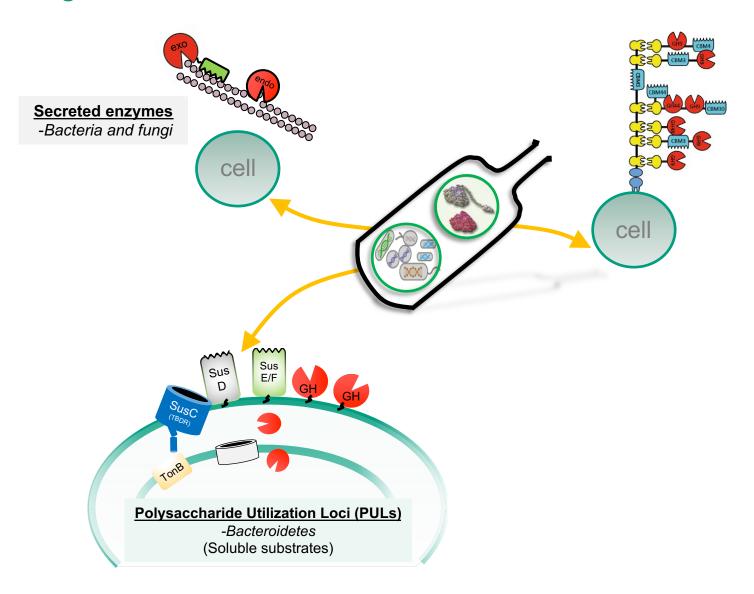
15+ families

Polysaccharide lyases (PLs)



Microbes use <u>similar</u> enzymes in <u>different</u> ways to degrade fiber

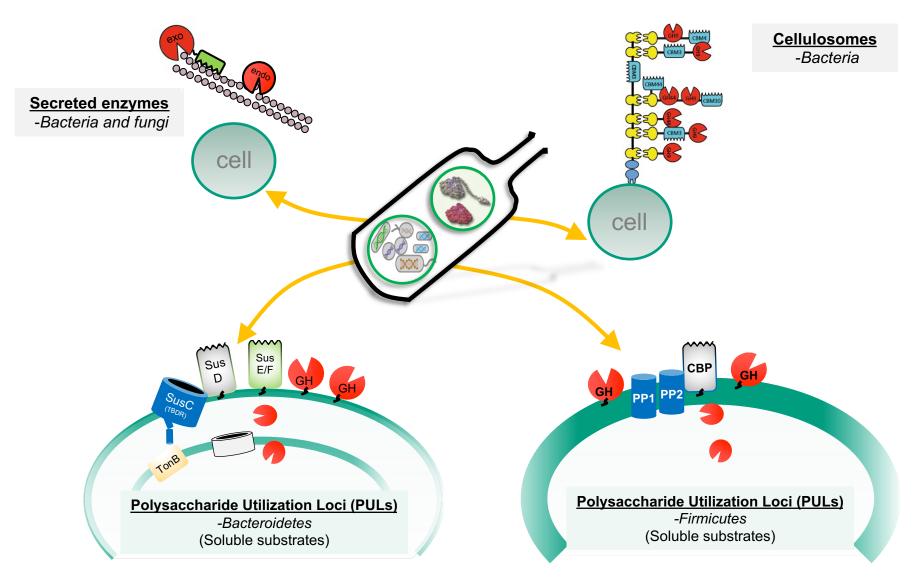




<u>Cellulosomes</u> -Bacteria

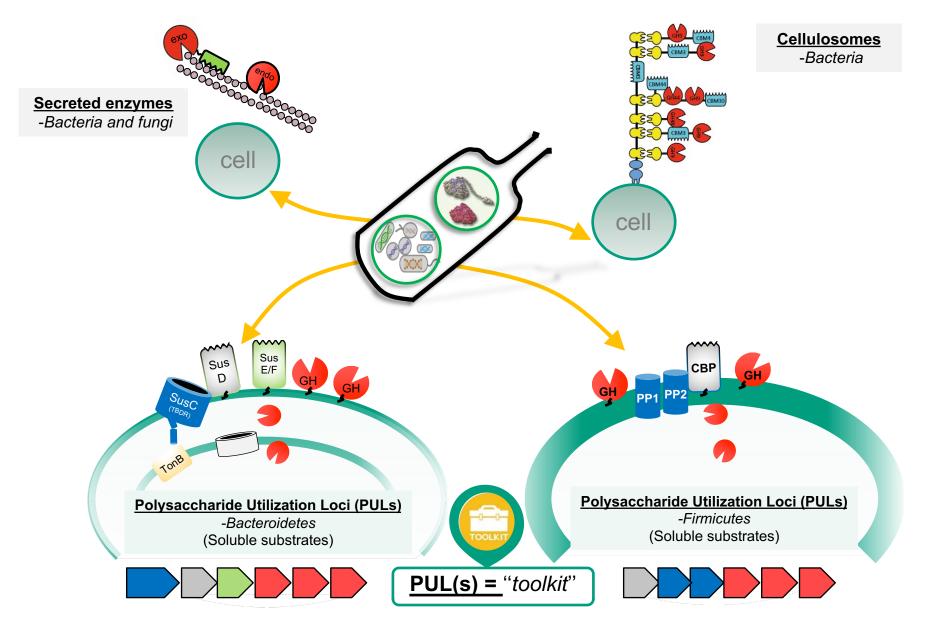
Microbes use <u>similar</u> enzymes in <u>different</u> ways to degrade fiber





Microbes use <u>similar</u> enzymes in <u>different</u> ways to degrade fiber





We combine *cultures and omics* to overcome the cultivability bottleneck **Metaproteomics** Metagenomics data Metatranscriptomic data Metaproteomic data Metabolomic data Metabolites identification and Metagenomics Metatranscriptomic data Peptide Identification and assembly and binning protein mapping metabolic model reconstruction mapping Vincent Eijsink **NMBU** MAGs: **M**etagenome **Enzymology A**ssembled Genomes Muller, et al. Trends in Microbiology, Vol 21

Meta-omics on digestive ecosystems

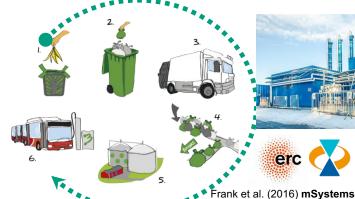


Pope et al. (2010) PNAS Vol 107 Pope et al. (2011) **ISME J**. Vol 5 Pope et al. (2011) Science Vol. 333



Naas, et al. (2014). mBio Vol 5 Arntzen et al (2017). Environ Microbiol, Vol 19 Naas et al. (2018). Microbiome, Vol 6

Hagen et al. (2021) ISME J.





Weimann et al. (2016) mSystems Hagen et al. (2017) **AEM** Kunath/Delogu et al. (2018) ISME J. Delogu et al. (2020) Nature Communications Jonassen et al. (2022) ISME J.



Rudi et al. (2017). AEM Vol 84 Minniti et al. (2017). Front Microbiol Vol 8 Minniti et al. (2019). Genes Vol 10



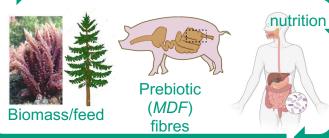
Pope et al. (2012) PLoS One

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Mackenzie et al. (2015) AEM Vol 81 Salgado-Flores et al. (2016) Micro. Gen. Solden et al. (2018) Nature Microbiology

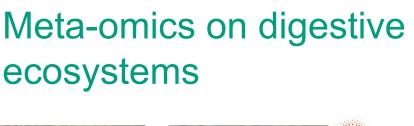


Taillefer et al. (2018) mSystems Emerson et al. (2018) Nature Microbiology



Omu et al. (2015). Microbiome La Rosa et al. (2018). Nature Communications Michalak et al. (2020). Nature Communications Linstad et al. (2021). mBio Ostrowski, et al. (2022) Nature Microbiology







Pope et al. (2010) **PNAS** Vol 107 Pope et al. (2011) **ISME J**. Vol 5 Pope et al. (2011) **Science** Vol. 333



Naas, et al. (2014). **mBio** Vol 5 Arntzen et al (2017). **Environ Microbiol**, Vol 19 Naas et al. (2018). **Microbiome**, Vol 6 Hagen et al. (2021) **ISME J.**

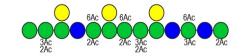


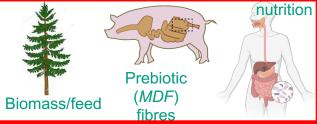


Rudi et al. (2017). **AEM** Vol 84 Minniti et al. (2017). **Front Microbiol** Vol 8 Minniti et al. (2019). **Genes** Vol 10



Pope et al. (2012) PLoS One
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Salgado-Flores et al. (2016) Micro. Gen.
Solden et al. (2018) Nature Microbiology





Omu et al. (2015). **Microbiome**La Rosa et al. (2018). **Nature Communications**Michalak et al. (2020). **Nature Communications**Linstad et al. (2021). **mBio**Ostrowski, et al. (2022) **Nature Microbiology**



Mannan degraders in the gut are *selfish*

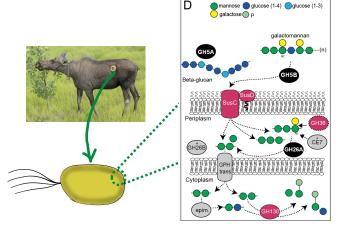




Interspecies cross-feeding orchestrates carbon degradation in the rumen ecosystem

Lindsey M. Solden', Adrian E. Naas², Simon Roux ¹, Rebecca A. Daly', William B. Collins³, Carrie D. Nicora⁴, Sam O. Purvine⁴, David W. Hoyt⁴, Julia Schückel³, Bodil Jørgensen², William Willats⁶, Donald E. Spalinger², Jeffrey L. Firkins⁵, Mary S. Lipton⁴, Matthew B. Sullivan ¹, Phillip B. Pope ² and Kelly C. Wrighton ¹

Solden et al. (2018) Nature Microbiology







Lindsey Solden Kelly Wrighton Ohio State University

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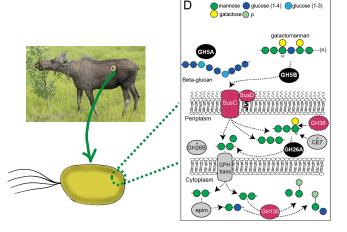




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Solden et al. (2018) Nature Microbiology





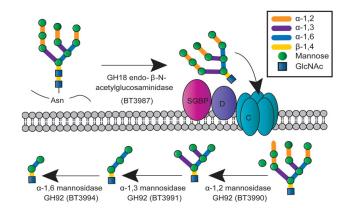


Lindsey Solden Kelly Wrighton Ohio State University

Human gut Bacteroidetes can utilize yeast mannan through a selfish mechanism

Fiona Cuskin, Elisabeth C. Lowe [...] Harry J. Gilbert

Nature 517, 165–169 (08 January 2015) | Download Citation ±



Mannan degraders in the gut are *selfish*

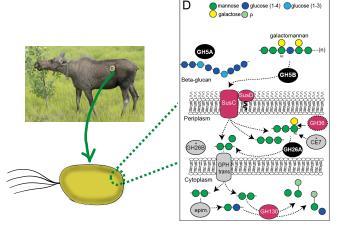




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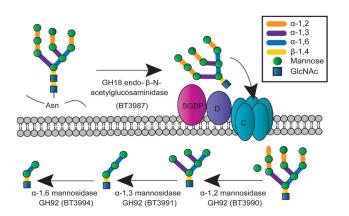


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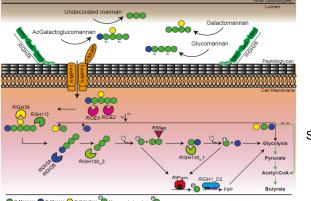
Nature 517, 165–169 (08 January 2015) | Download Citation ±





The human gut Firmicute Roseburia intestinalis is a primary degrader of dietary β -mannans

Sabina Leanti La Rosa 🐧 Maria Louise Leth², Leszek Michalak¹, Morten Ejby Hansen 🐧 ², Nicholas A. Pudlo³,





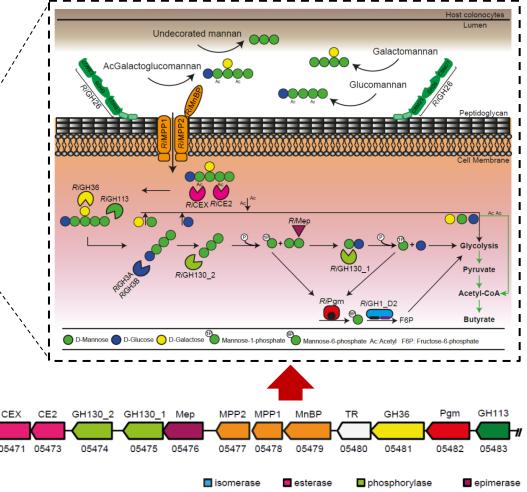
Sabina La Rosa NMBU

Roseburia intestinalis is a primary degrader of mannan





- 2. Needs CEs
- 3. It is "selfish"
- It produces butyrate
- 5. Associated with improved gut health and low intestinal disease states



Mannan PUL a.k.a. "Mannan toolkit"



GH₁

05469/70

□ isomerase □ esterase □ phosphorylase □ epimerase □ ABC transporter □ transcriptional regulator □ α-galactosidase □ mutase □ mannanase/mannosidase □ glucosidase

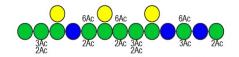
La Rosa et al. (2019) Nature Communications

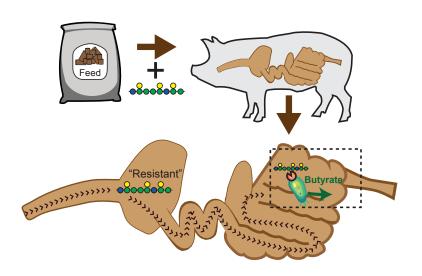
Can we take advantage of the unique features of mannan and its degraders?





Mannan as a prebiotic?

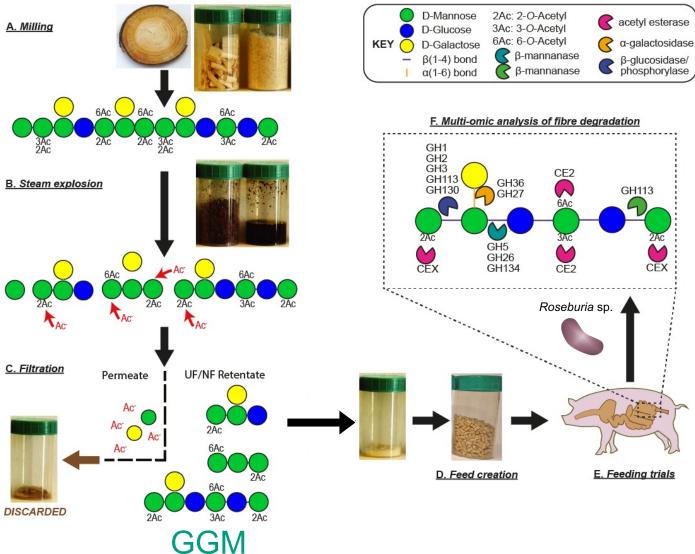




- The (few) mannan degraders characterized thus far seem to be "selfish" (i.e. selective)
- 2. Mannan substrates requires a particular set of CAZymes (often GHs, CEs and CBMs) that are not widely found in gut commensals (*i.e.* selective)
- 3. Several famous butyrate-producing commensal bacteria are believed to degrade mannan (*i.e.* Roseburia sp.)

Mannan from wood.....and into feed?







Bjørge Westereng NMBU



Leszek Michalak NMBU



J. Chris Gaby NMBU



Sabina La Rosa NMBU

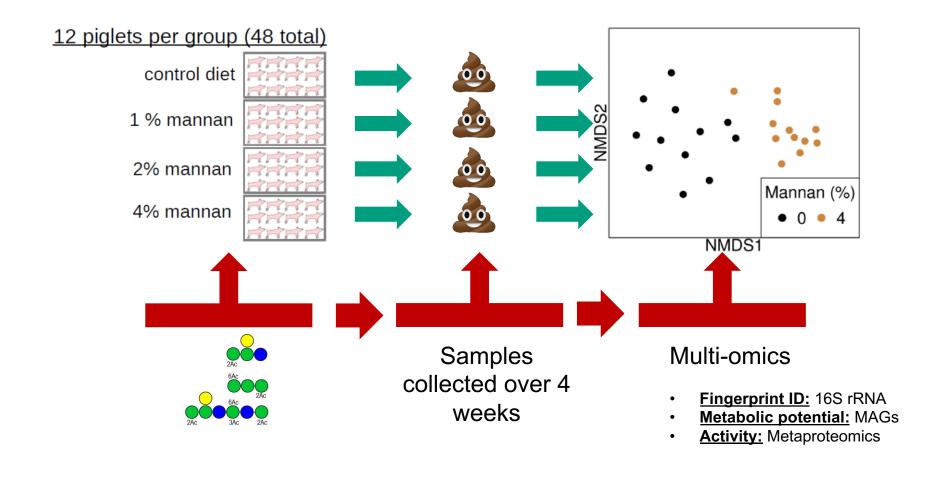


Leidy Lagos NMBU

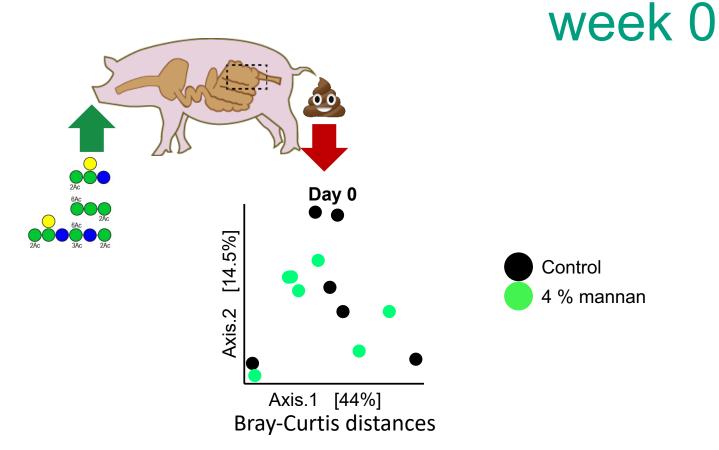
La Rosa et al. (2019) Nature Communications



Dietary mannan inclusion causes a dose response in the gut microbiome

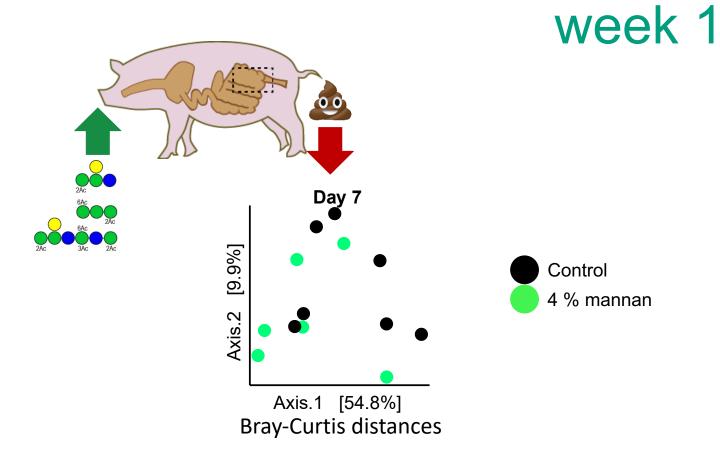






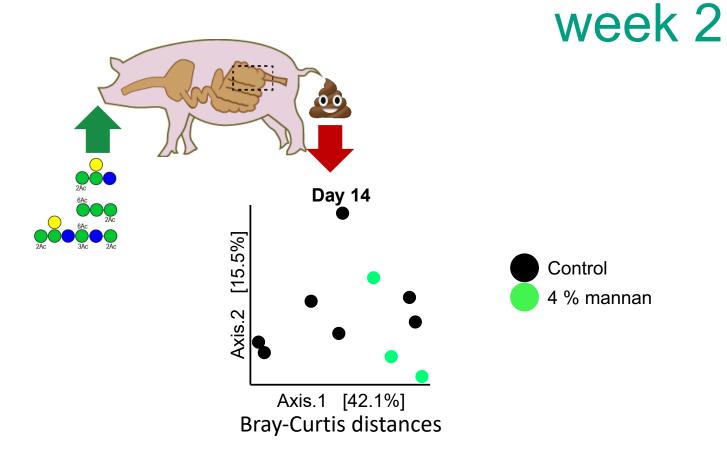






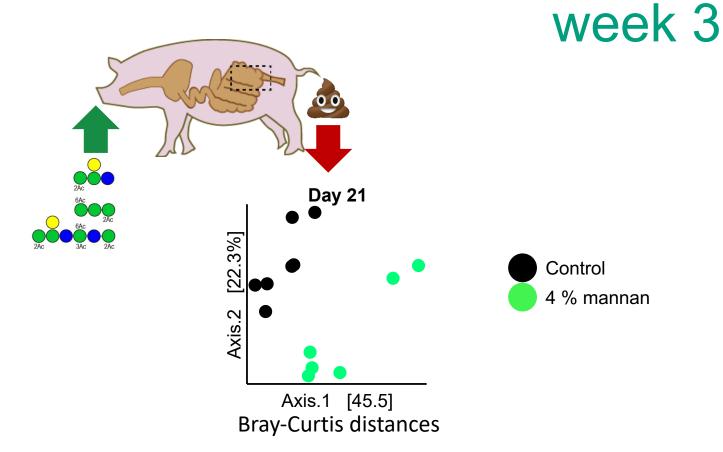






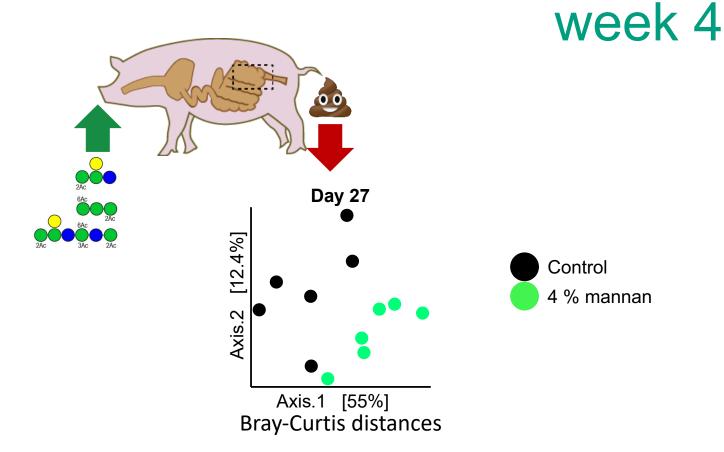






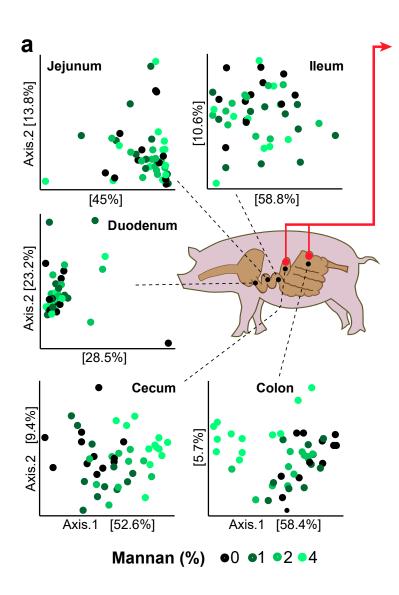








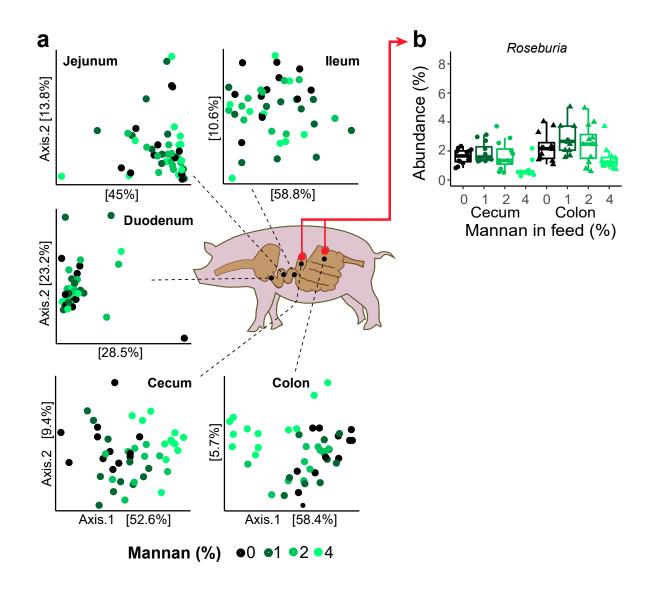




Mannan is degraded in the **distal gut**

The amount of mannan added seemingly has an effect on key commensals

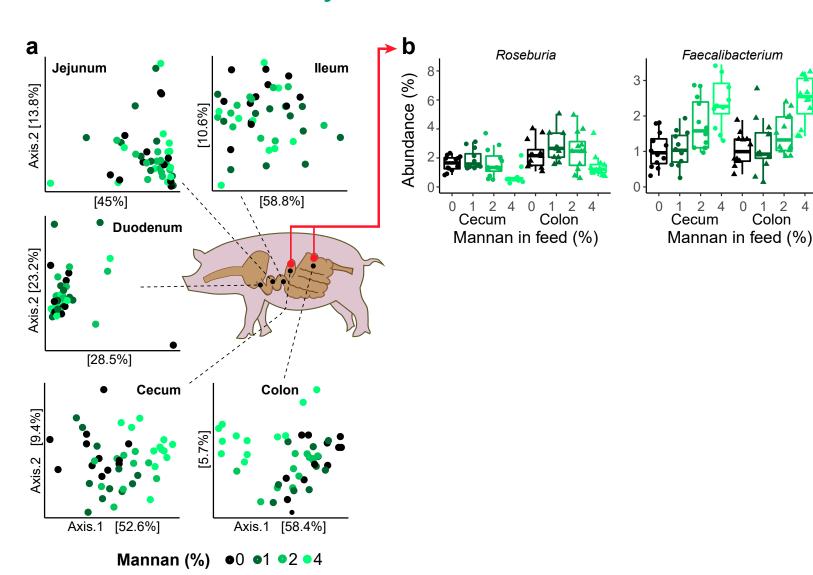




The amount of mannan added seemingly has an effect on key commensals

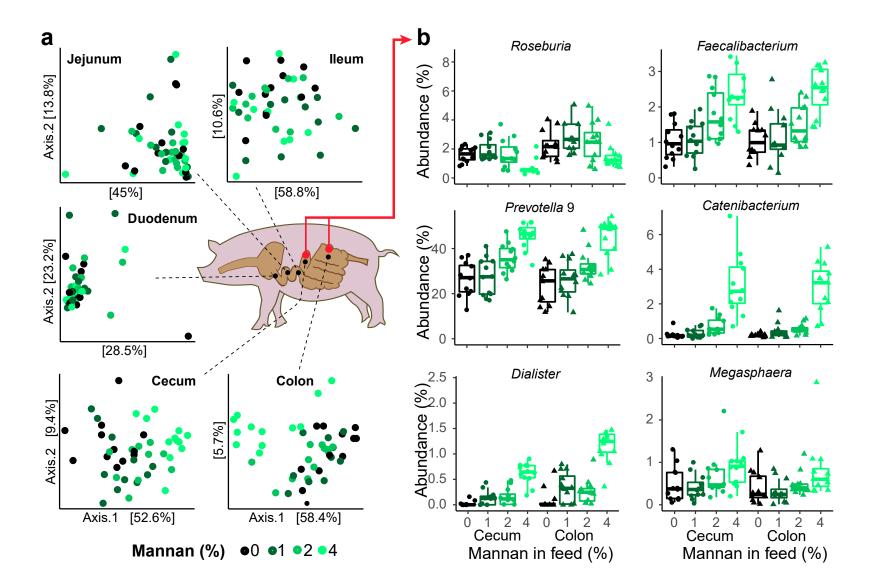


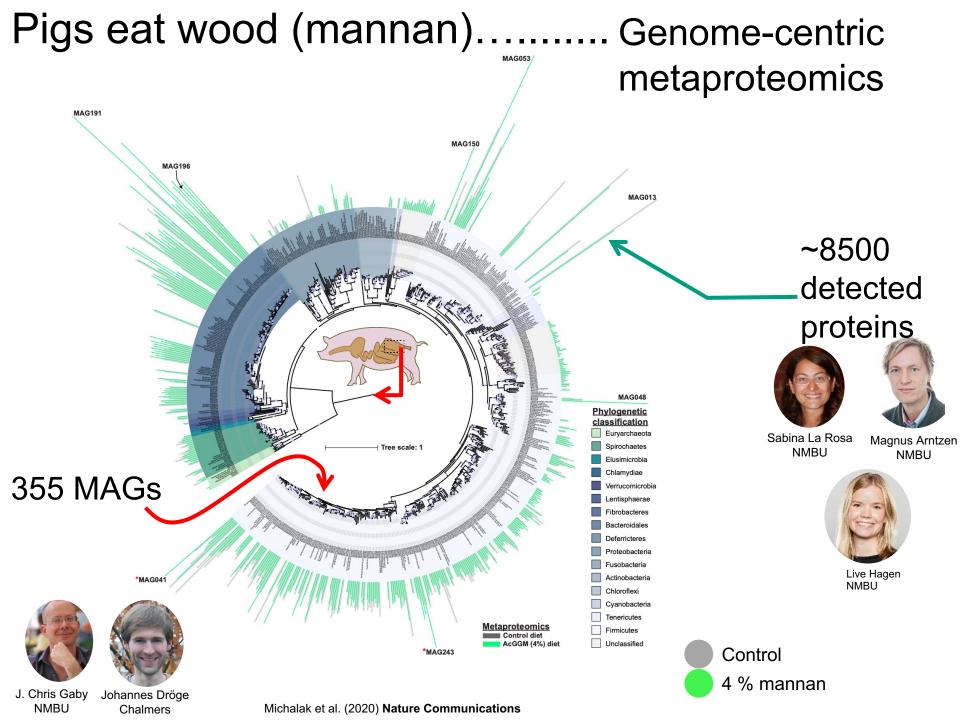
Colon

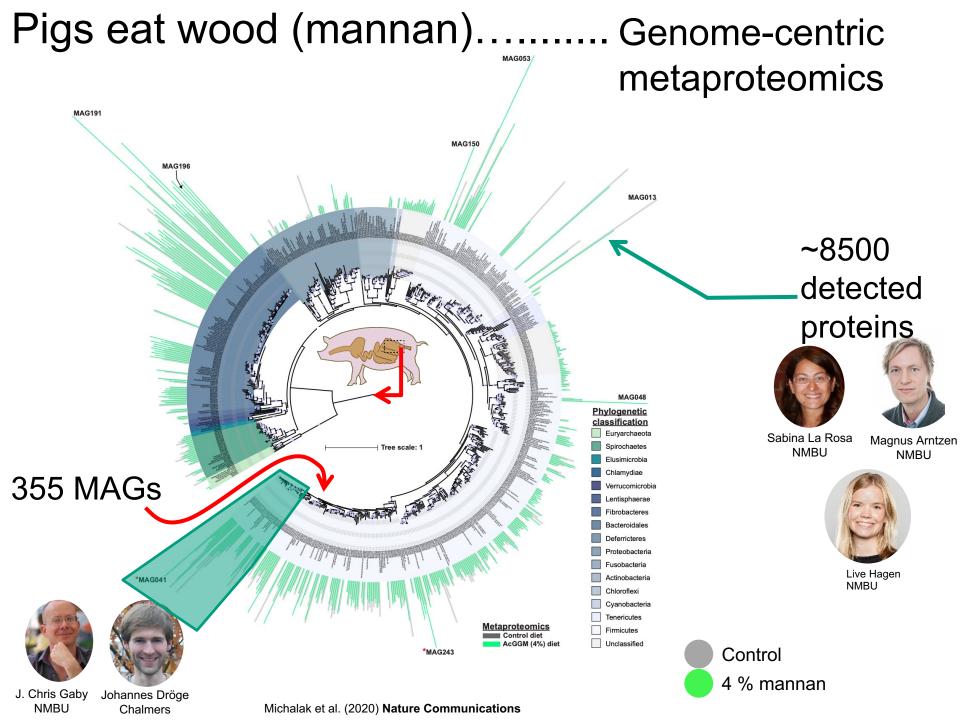


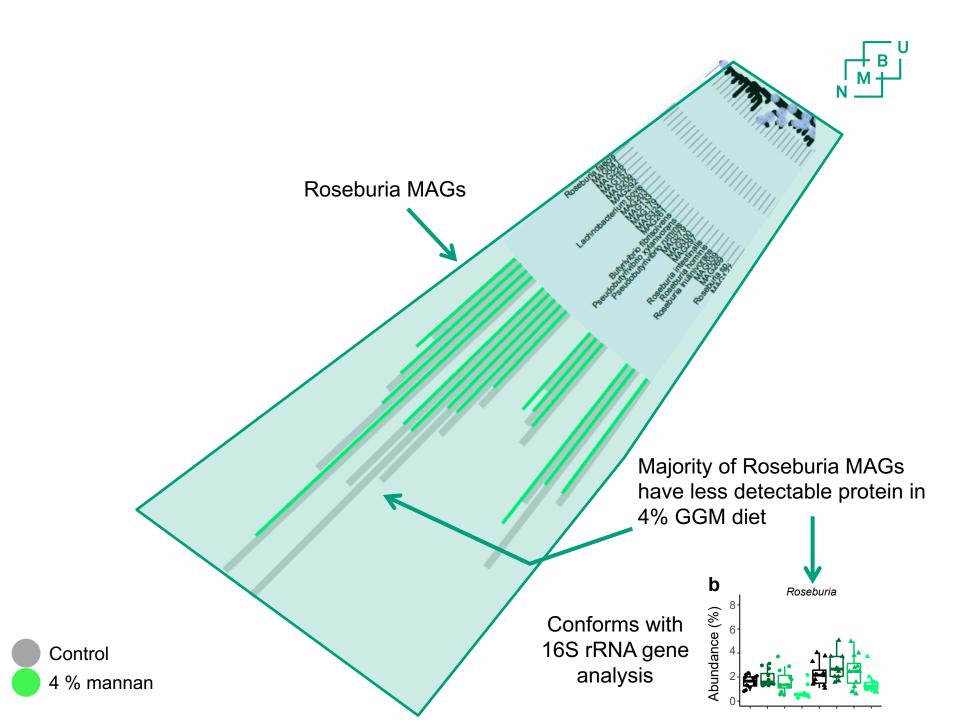
The amount of mannan added seemingly has an effect on key commensals

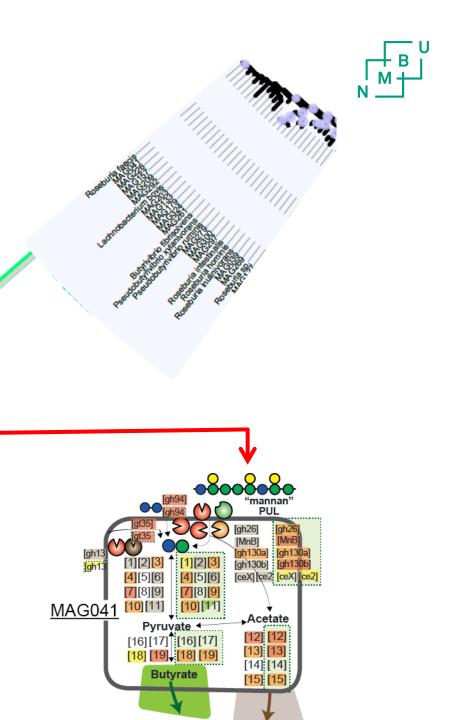












EXCEPT for:

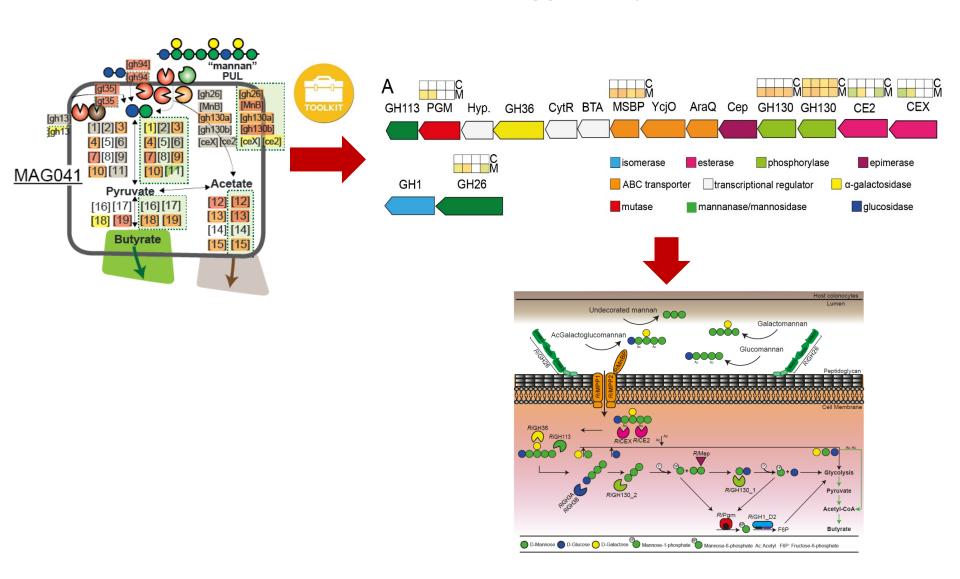
*MAG041

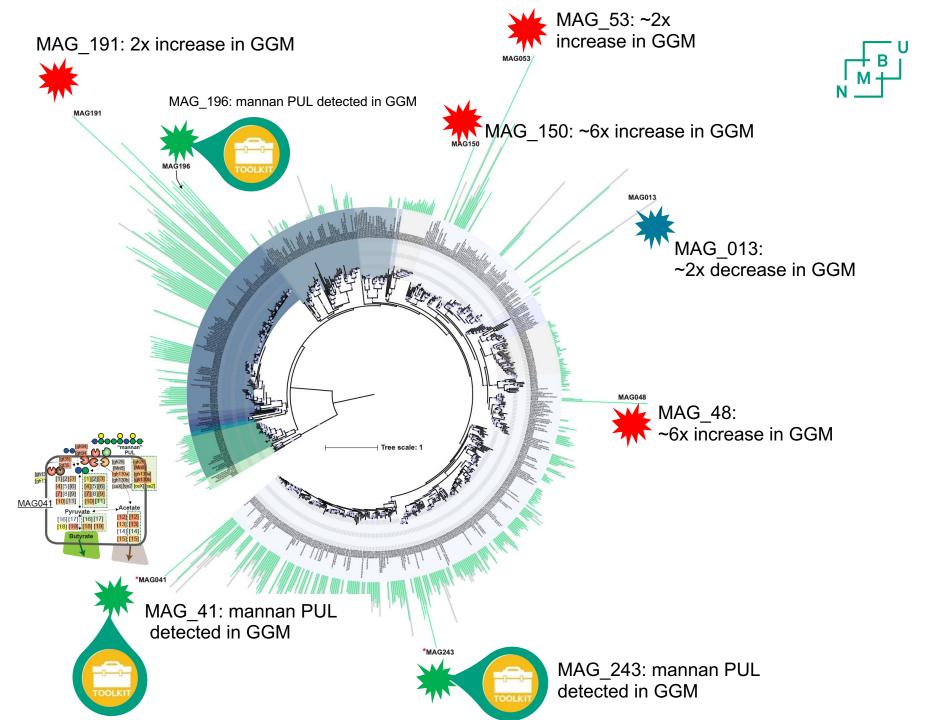
Which also encodes a mannan PUL that is detected in GGM

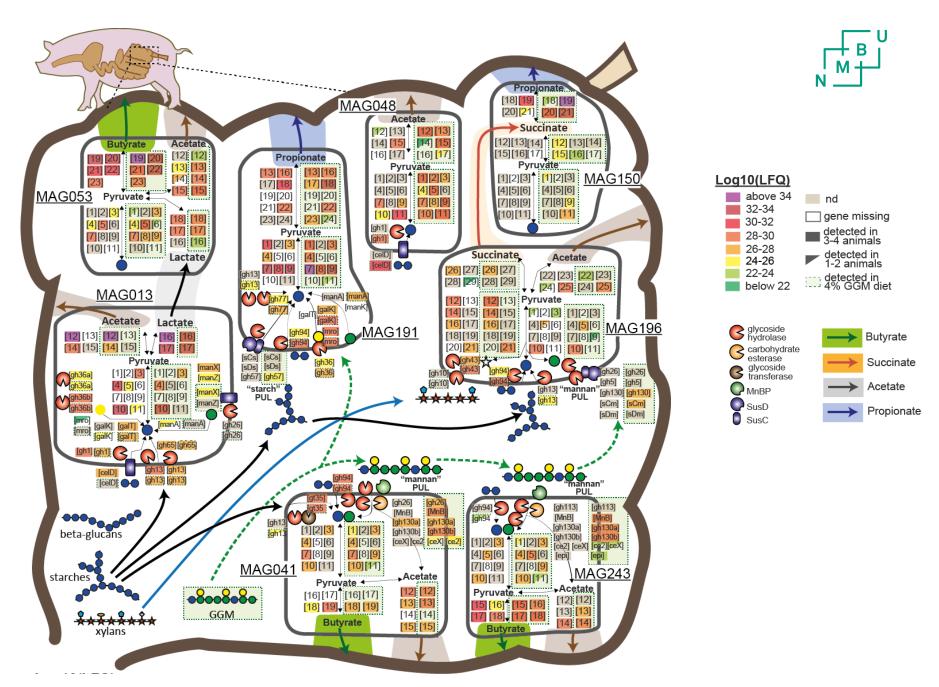
Control
4 % mannan

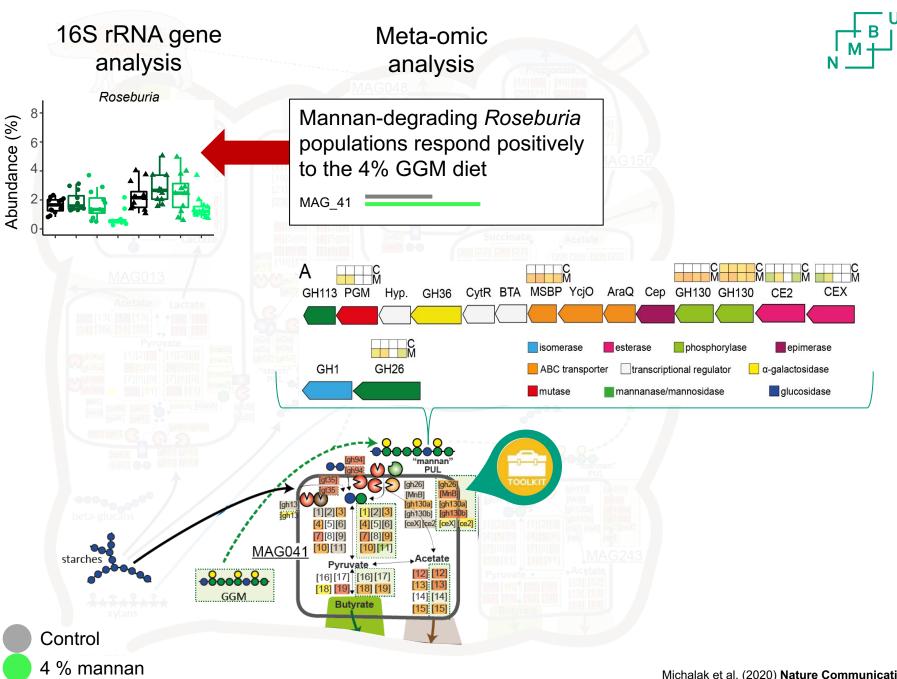


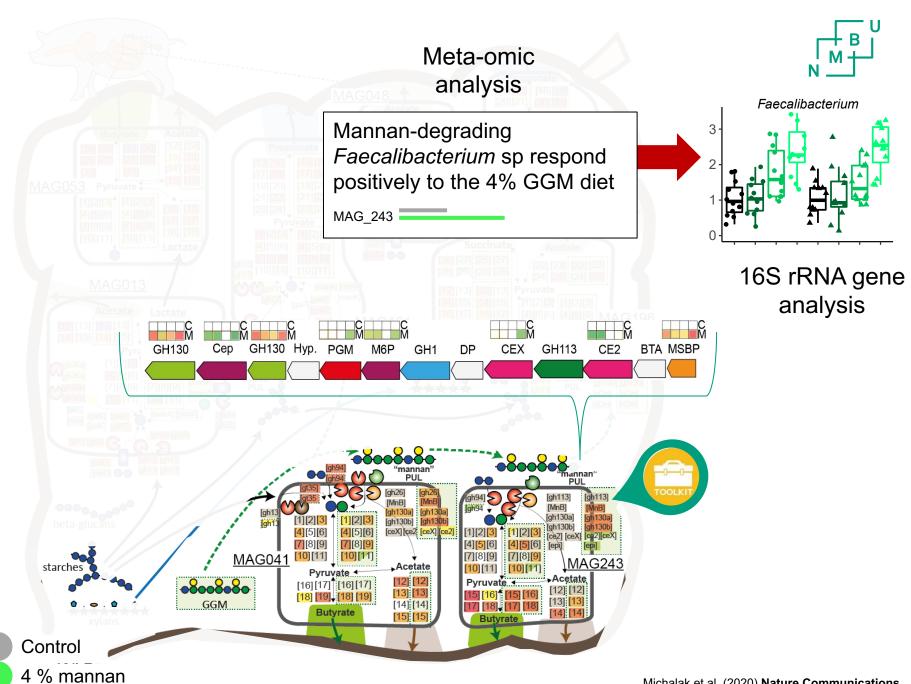
MAG_41 has a "selfish" mannan PUL triggered by AcGGM

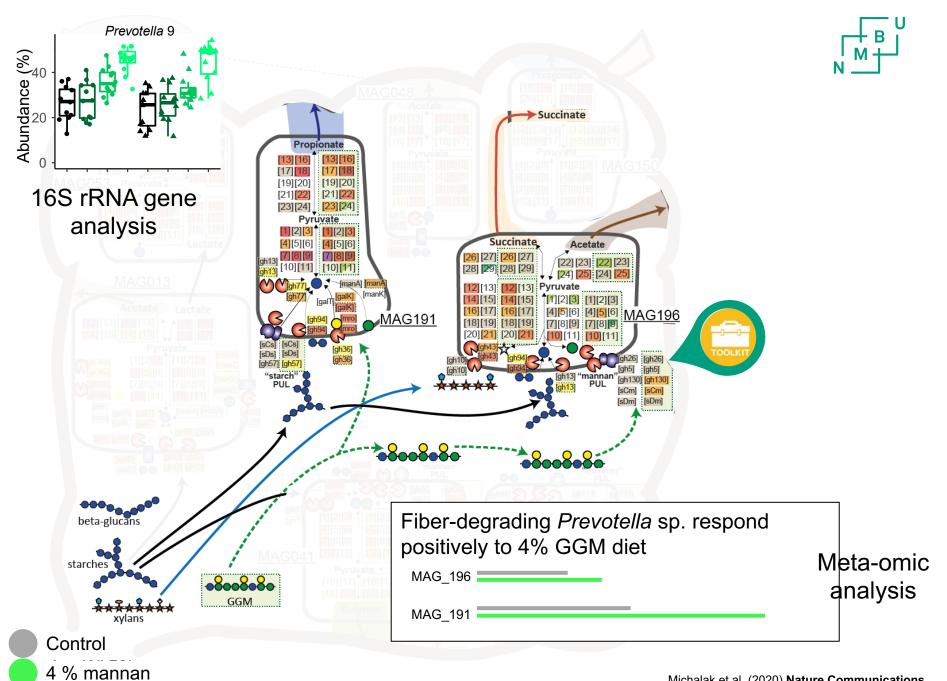




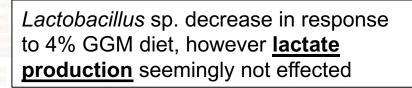












Meta-omic analysis

[galK] [galT] [manA] [manA] [gh26] [gh26]

MAG_013

Control

starches

4 % mannan

beta-glucans

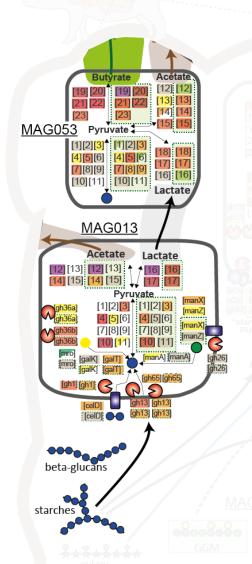
MAG013

Acetate

12 [13] [12] [13] 16 [16] [16] [17] [17]

Lactate





Megasphaera sp. (<u>lactate utilizers</u>) increase

MAG_53



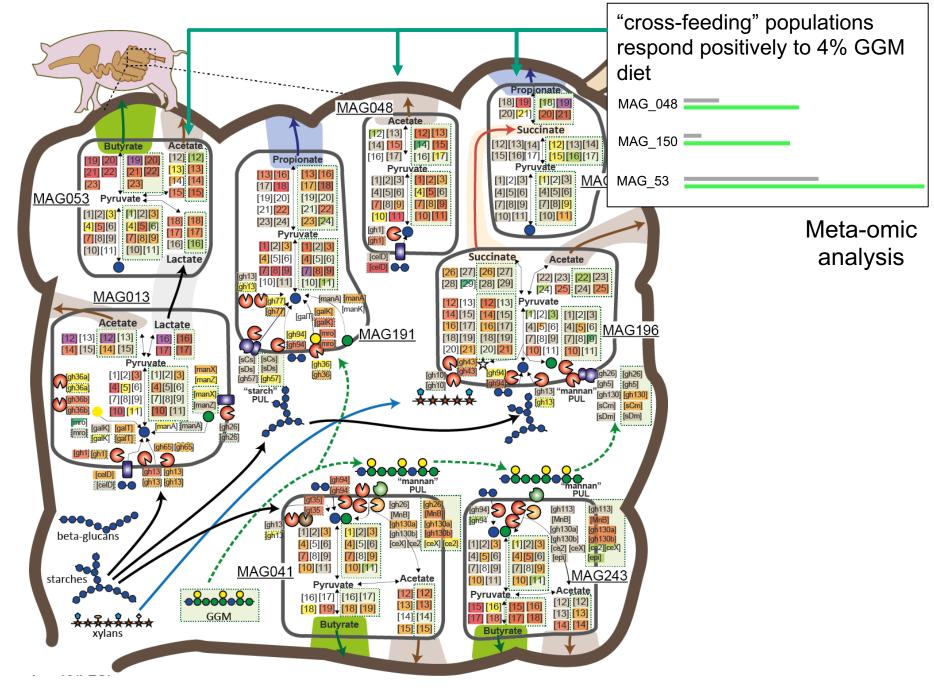
Lactobacillus sp. decrease in response to 4% GGM diet, however <u>lactate</u> <u>production</u> seemingly not effected

MAG_013

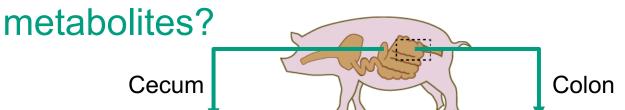
Meta-omic analysis

Control

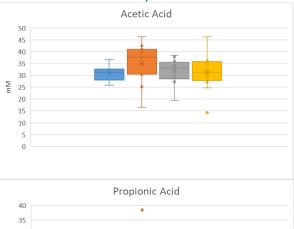
4 % mannan

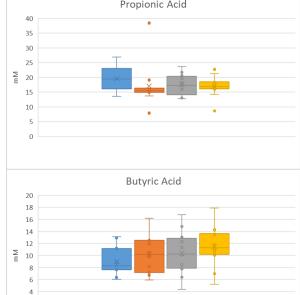


What effect does AcGGM have on

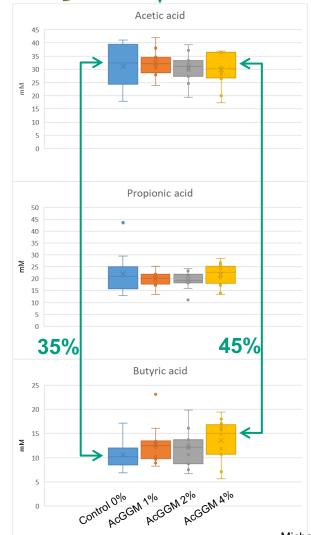








Countal Dolo Aceem Ace



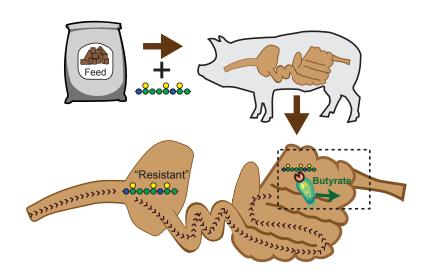
Acetate:propionate:butyrate (ratio)

Control = 100 : 72 : 35 AcGGM (4%) = 100 : 74 : 45

p-value = 0.004

So what does this all tell us?

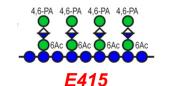




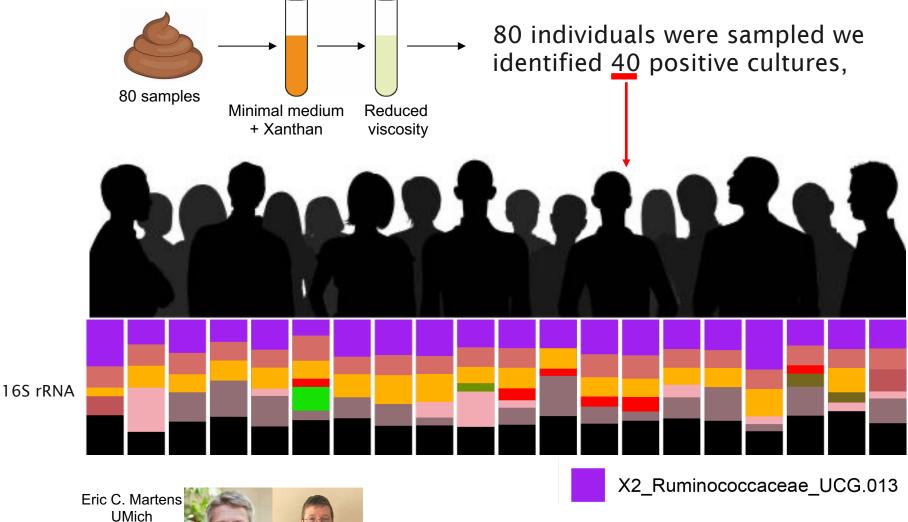
Mannan as a prebiotic?

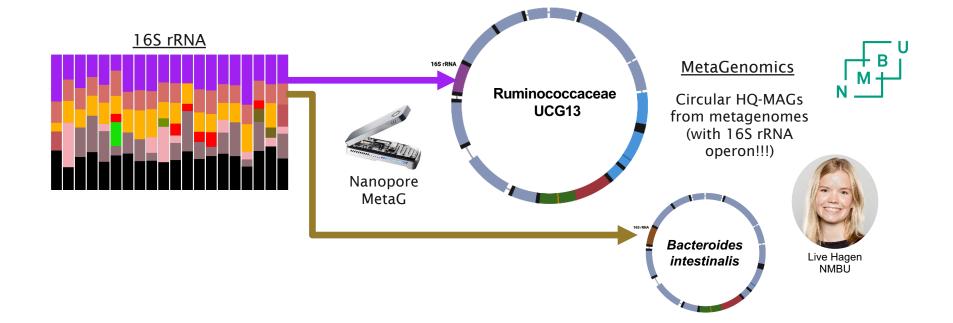
- 1. Understand your microbes and their tools
- Understand your <u>substrate</u>
- 3. Even <u>if you can manage to</u> "tap your specific microbe on the head", there will be a "butterfly effect". To understand this requires "more of everything"

We eat MDFs every day



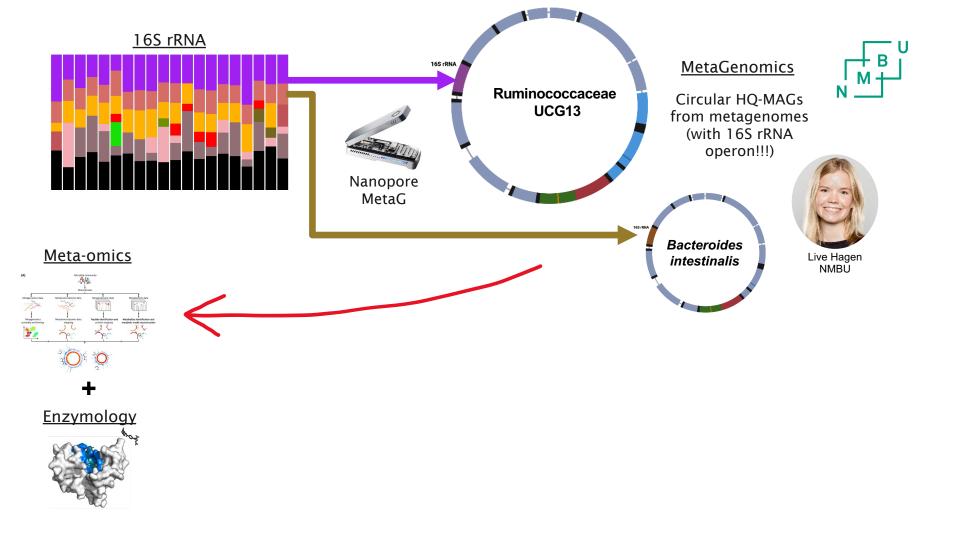










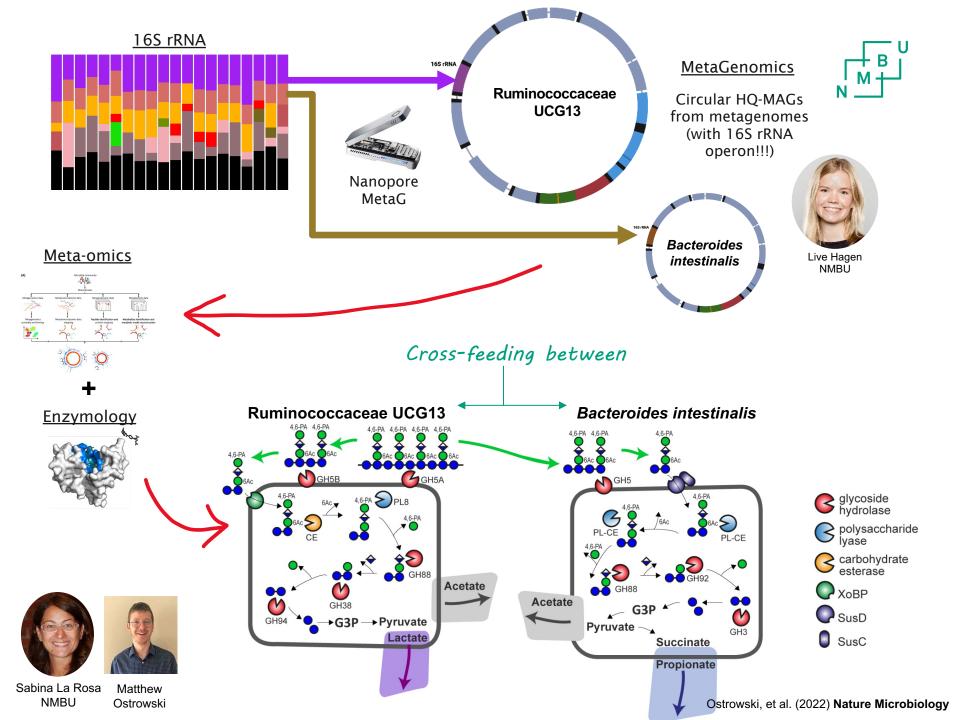




NMBU

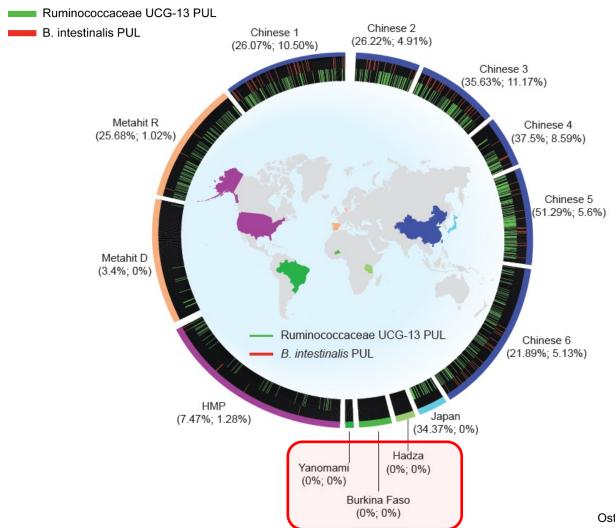


Ostrowski



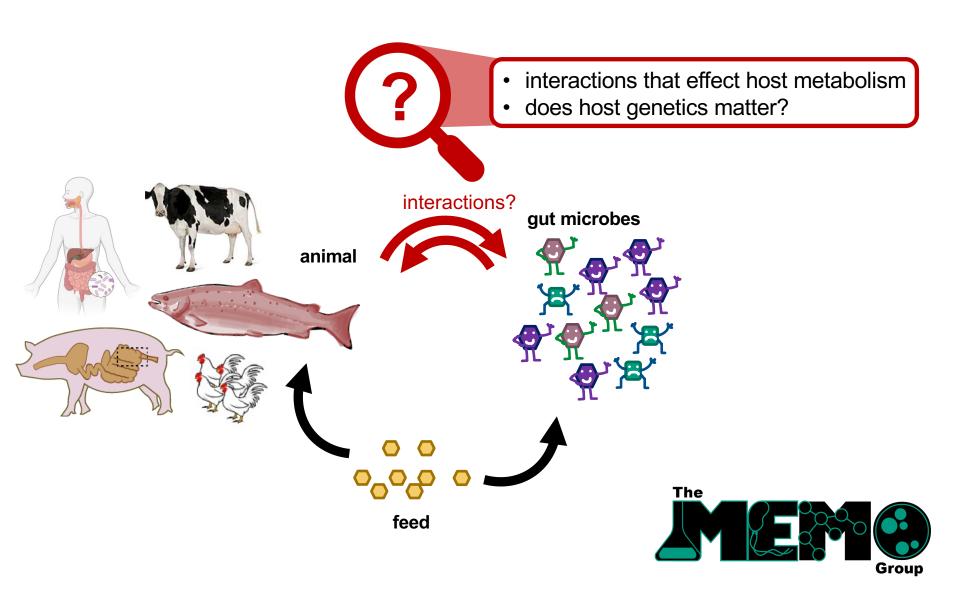
Xanthan utilization loci are widespread in modern microbiomes





So, where are we going from here?



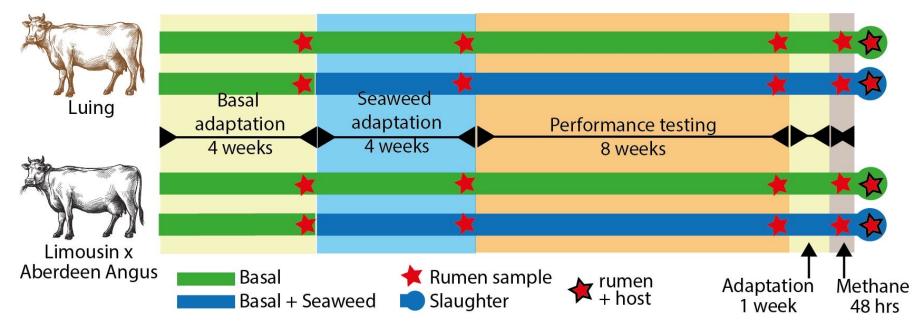




SuPAcow:



The feed-microbiome-host axis in cows



Thea Os Andersen



lanina (Yanna) Altshuler



Torgeir Hvidsten



Rainer Roehe



Carl Kobel

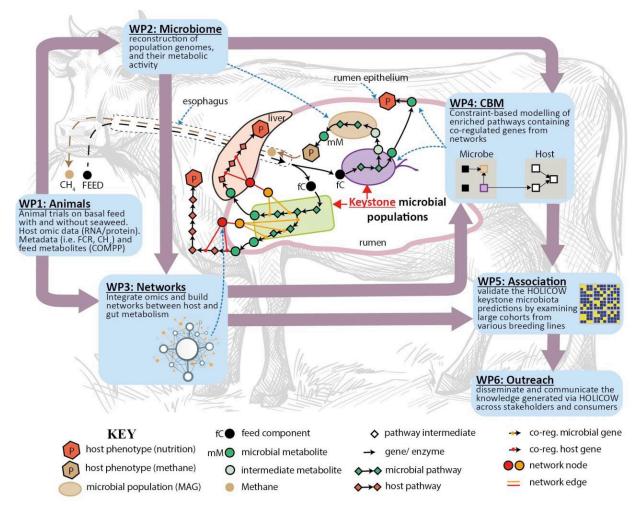


novo nordisk fonden



SuPAcow: The feed-microbiome-host axis in cows





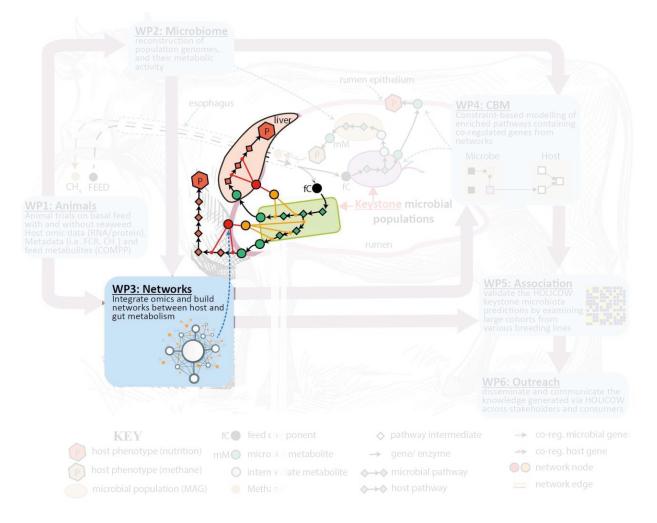
novo nordisk fonden



SuPAcow:



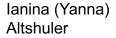
The feed-microbiome-host axis in cows





Do proteins expression levels interact across species?

> keystone protein groups detected in at least 12 of 32 of (control) animals



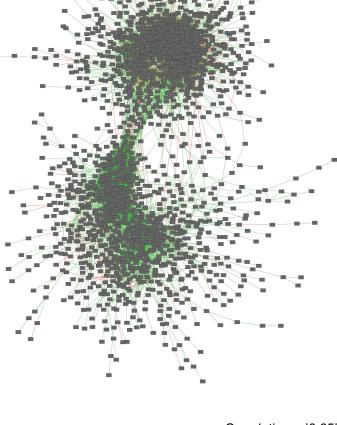


Rainer Roehe



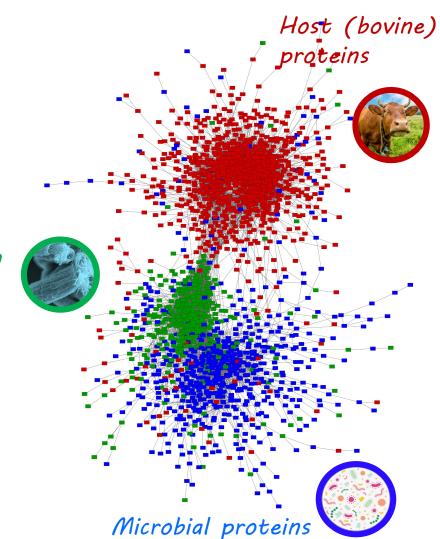
novo nordisk fonden







Do proteins expression levels interact across species?



Entodinium caudatum (eukaryote) proteins



Identification of modules with WGCNA analysis

Groups of proteins that share similar expression pattern

that are consistent across individual cows

Identified 9 modules



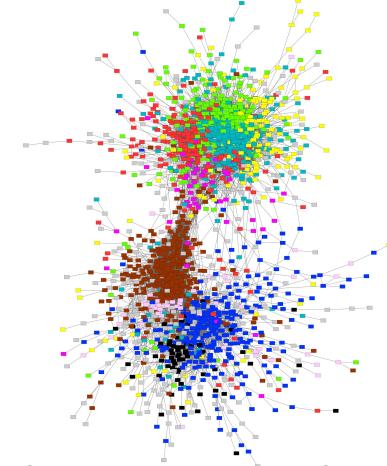
5 bovine



2 protozoan

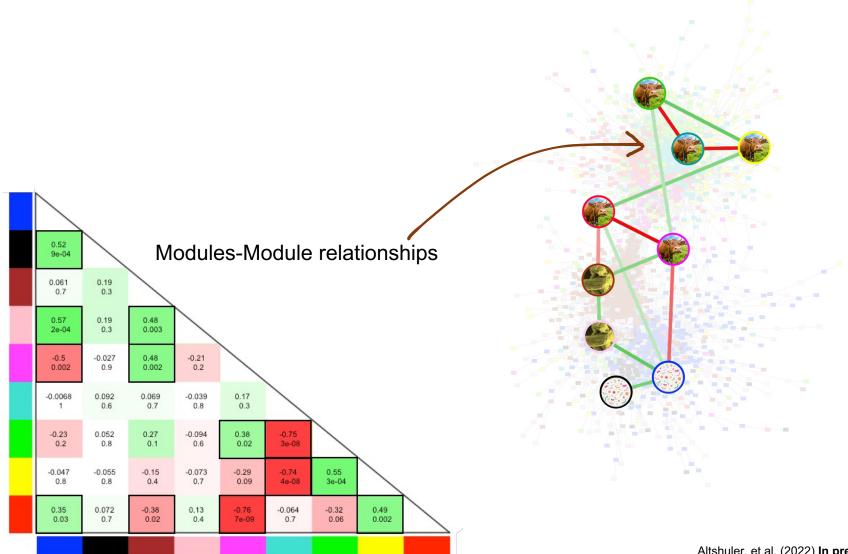


2 microbial

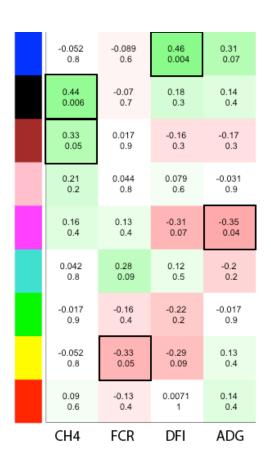


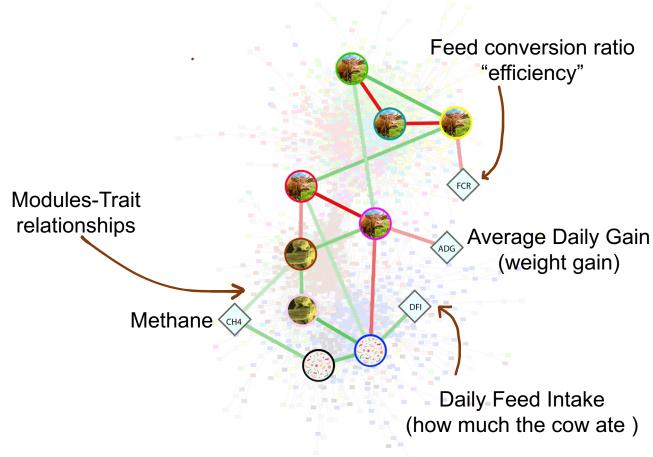
Weighted Gene Co-expression Network Analysis (WGCNA)



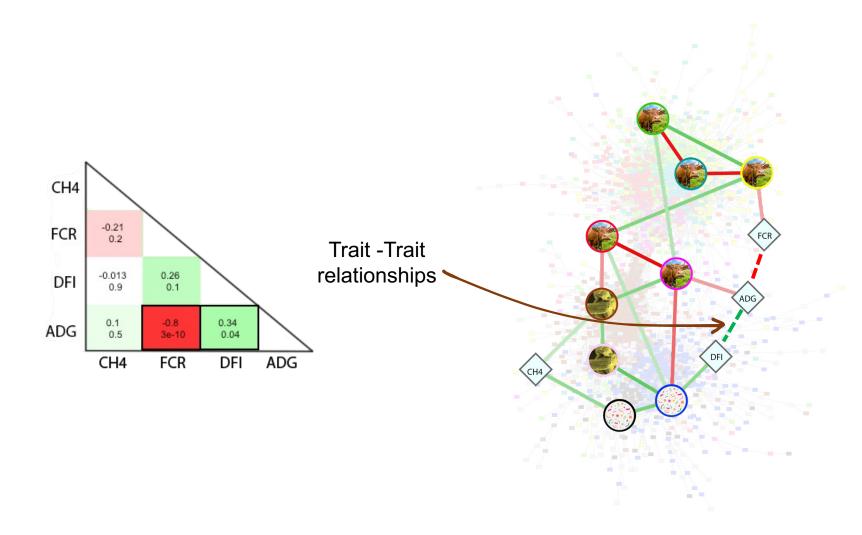












Enrichment analysis of modules



Enrichment of organisms



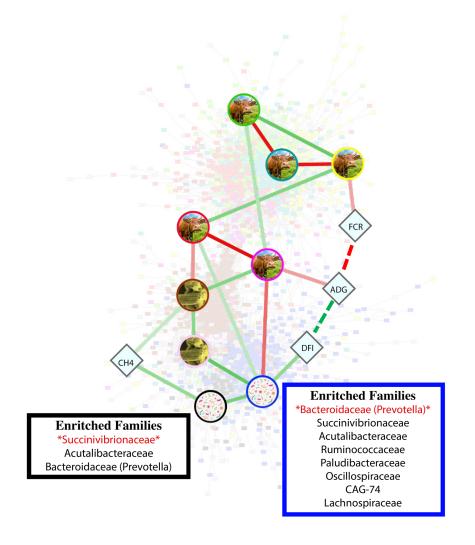
5 bovine



2 protozoan

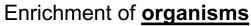


2 microbial



Enrichment analysis of modules







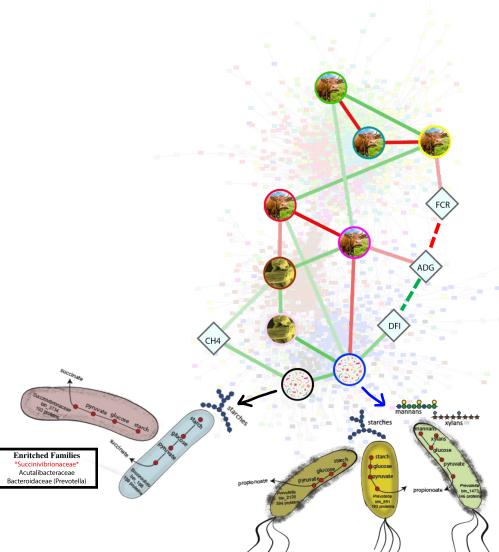
5 bovine



2 protozoan



2 microbial



Enritched Families *Bacteroidaceae (Prevotella)*

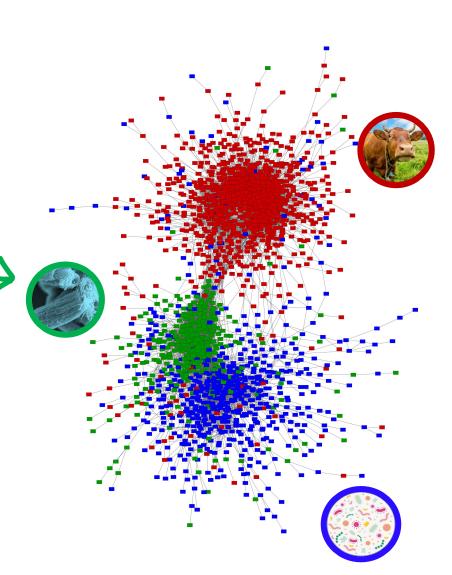
Succinivibrionaceae Acutalibacteraceae Ruminococcaceae Paludibacteraceae Oscillospiraceae CAG-74 Lachnospiraceae

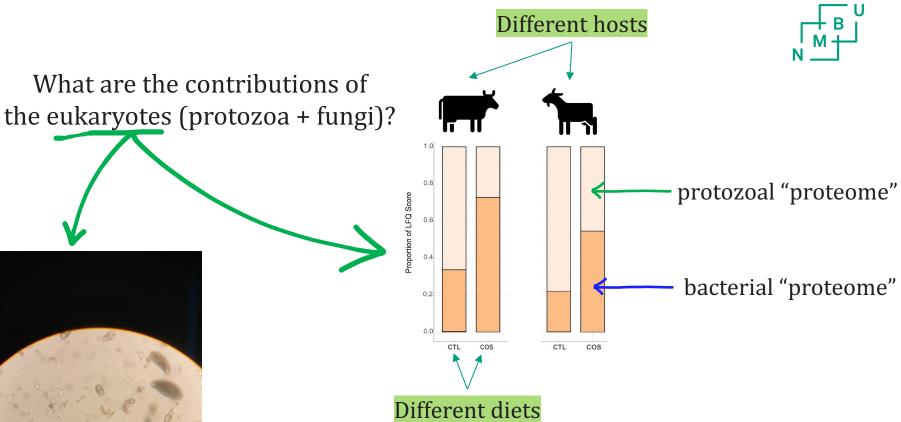


What are the contributions of the eukaryotes (protozoa + fungi)?

Prokaryotes	50-90%
Protozoa	10-50 %
Fungi	5-10%

% of microbial biomass in the rumen



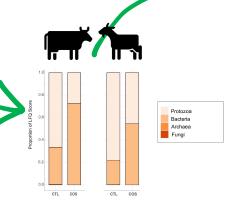




Thea Os Andersen

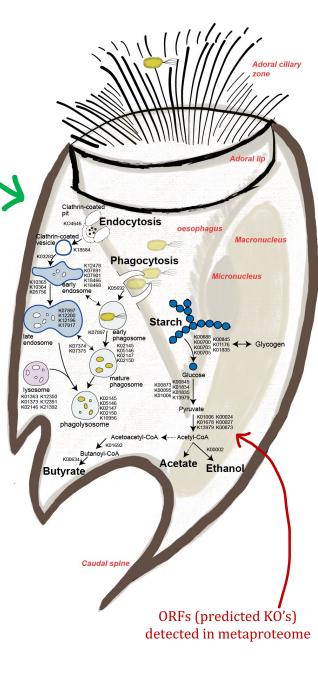


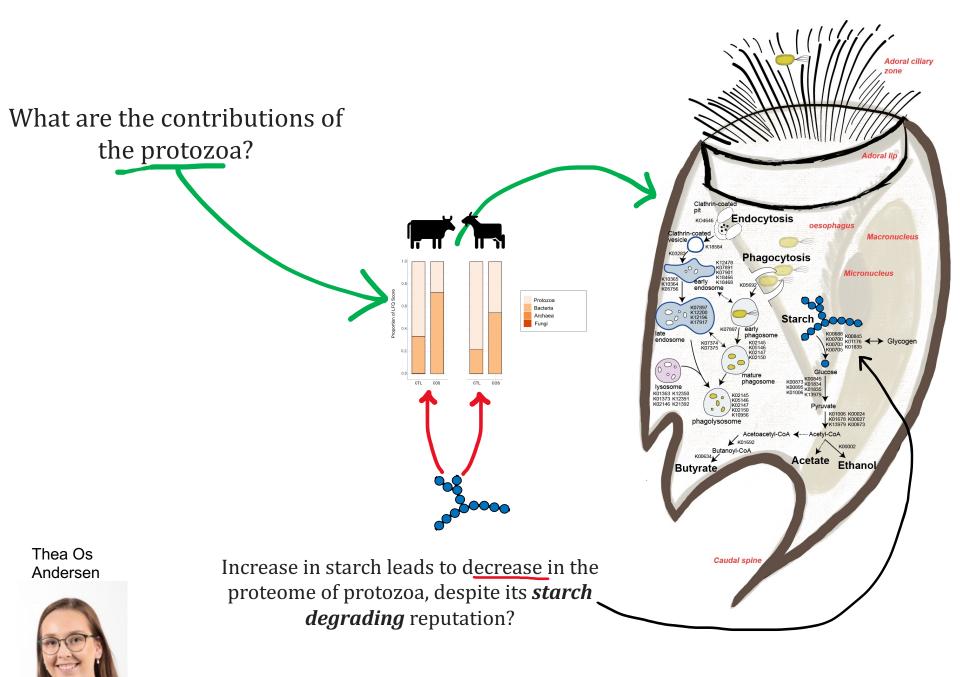
What are the contributions of the protozoa?



Thea Os Andersen



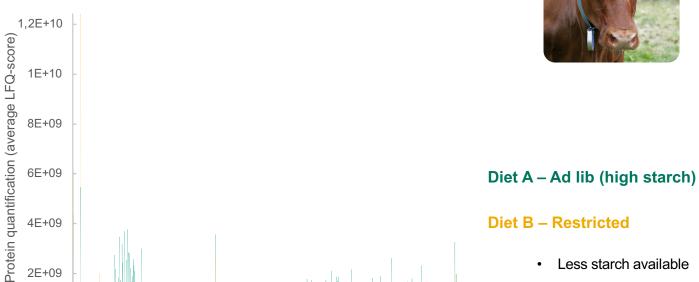








2E+09



Eukaryota genomes and prokaryota Metagenome-Assembled Genomes (MAGs)



Magnus Ø. Arntzen **NMBU**

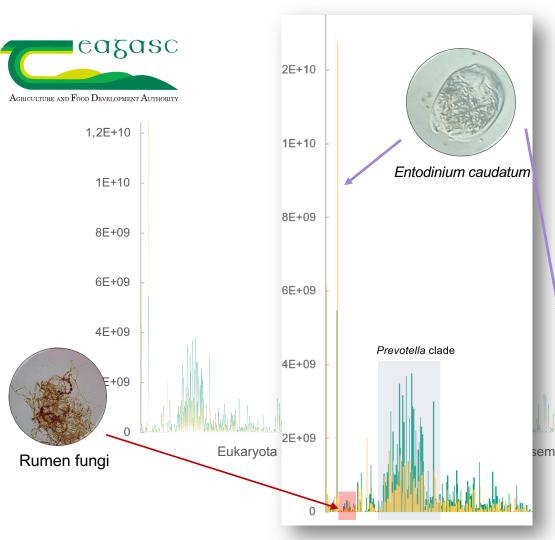


Live H. Hagen **NMBU**

Norwegian University of Life Sciences

Less starch available







Diet A – Ad lib (high starch)

Diet B - Restricted

Less starch available



sembled Genomes (MAGs)



Magnus Ø. Arntzen NMBU



Live H. Hagen NMBU

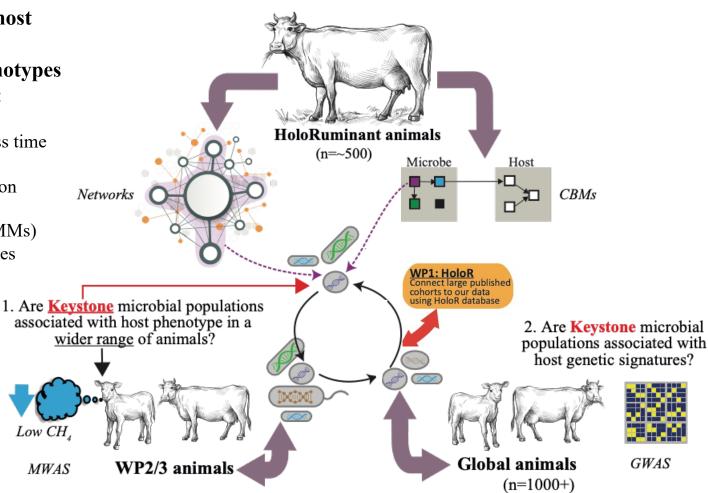
Norwegian University of Life Sciences





Genetic variation in the host will be connected to microbiome-related phenotypes by using a combination of:

- microbiome dynamics across time and its potential sources
- metagenome-wide association studies (MWAS)
- bayesian mixed models (BMMs)
- existing public GWAS studies performed on a global level



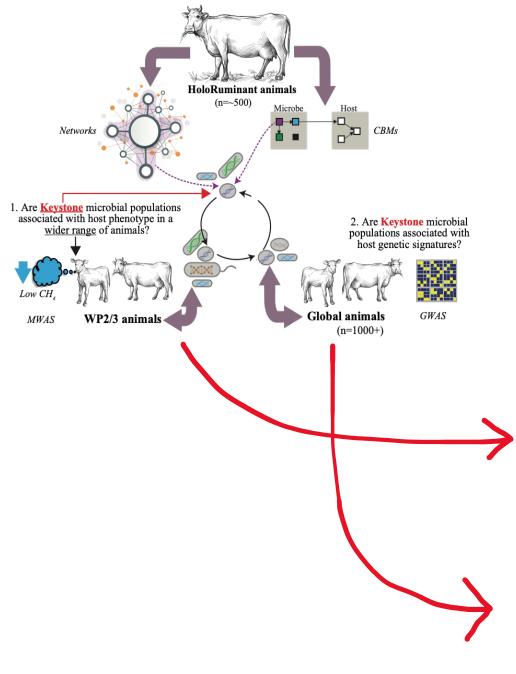




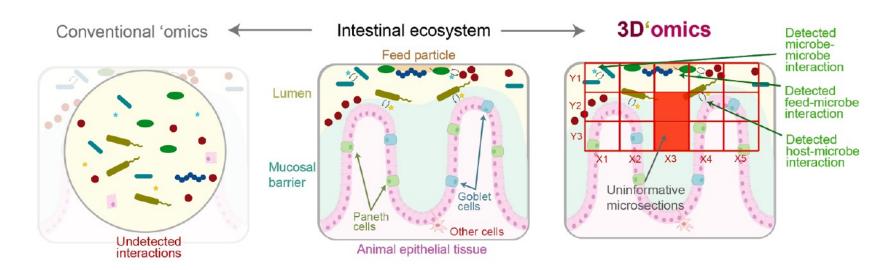
Table: List of data that will be stored in HoloR for use by WP4

Project/Origin	Breed	#animals	Genotyped	Phenotyped	MetaTaxo	
						omics
HoloRuminant	Holstein (calf)	500+	⋖	⋖	⋖	⋖
	Holstein-Friesian (calf)	180+	⋖	⋖	⋖	⋖
	Norwegian Red (calf)	40	⋖	⋖	⋖	⋖
	Finnish Nordic Red (calf)	40	×	⋖	⋖	×
	Charolais (calf)	40	⋖	⋖	⋖	⋖
	Jersey (calf)	30	⋖	⋖	⋖	⋖
	Normande (calf)	30	⋖	⋖	⋖	⋖
	Angus (calf)	20	⋖	⋖	⋖	⋖
	Murciano-Granadina (goats)	100	⋖	⋖	⋖	⋖
	Romane (sheep)	100	×	⋖	⋖	⋖
	Laucane (sheep)	50	⋖	⋖	⋖	⋖
	Segureña (sheep)	50	⋖	⋖	⋖	⋖
Ruminomics	Holstein (calf)	816	⋖	⋖	⋖	×
	Finnish and Swedish Nordic Red (cows)	200	⋖	⋖	⋖	×
CSIRO	Brahman (calf)	56	⋖	⋖	⋖	⋖
MASTER/TEA GASC	Limousin	500	⋖	⋖	⋖	≪ *
INRAE	Holstein	108	⋖	⋖	⋖	≪ *
	Charolais	61	⋖	⋖	⋖	×
AGR	Romney and Coopworth mix (sheep)	1100	⋖	⋖	⋖	×
SRUC	Aberdeen Angus	104	⋖	⋖	⋖	⋖
	Limousin	103	⋖	⋖	⋖	⋖
	Charolais	71	⋖	⋖	⋖	⋖
	Luing	85	⋖	⋖	⋖	⋖
UAL	Angus	200	⋖	⋖	⋖	⋖
	Charolais	150	⋖	⋖	⋖	⋖
	Hybrid (UAL)	500	⋖	⋖	⋖	⋖





"Three-dimensional holo'omic landscapes to unveil host-microbiota interactions shaping animal production"



Antton Alberdi



Phil B. Pope

Torgeir Hvidsten

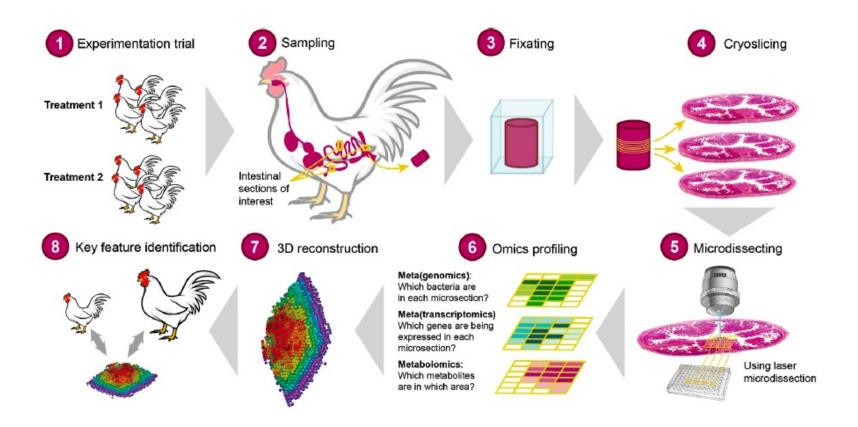


Irma Oskam Bjørge Westereng











> WE ARE HIRING!





MEMO scientists

- Sabina La Rosa
- Live Hagen
- Arturo Vera Ponce De Leon
- Adrian Naas
- Magnus Ø. Arntzen
- Thea Os Andersen
- Ianina Altshuler
- Alessandra Ferrillo
- Carl M. Kobel
- Valerie Schiml
- Juline Walter













@ThePopeLab

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Some images created with BioRender.com









